

1 **Evaluation of QuantiFERON®-TB Gold-Plus in Healthcare Workers in a Low-**
2 **Incidence Setting**

3

4 Hee-Won Moon,^{1,3} Rajiv L. Gaur¹, Sara Shu-Hwa Tien⁴, Mary Spangler⁴, Madhukar Pai⁵ and
5 Niaz Banaei^{1,2,6}

6 ¹Pathology and ²Medicine, Division of Infectious Diseases & Geographic Medicine, Stanford
7 University School of Medicine, Stanford, CA, ³Department of Laboratory Medicine, Konkuk
8 University School of Medicine, Seoul, Korea, ⁴Occupational Health Clinic, Stanford Health
9 Care, ⁵Department of Epidemiology, Biostatistics and Occupational Health, McGill
10 University, Montreal, Canada, ⁶Clinical Microbiology Laboratory, Stanford Health Care, CA,
11 USA

12

13

14 **Running title:** Evaluation of QuantiFERON-TB Gold-Plus

15

16

17

18 **Correspondence**

19 Niaz Banaei, MD

20 Rm. 1602, 3375 Hillview Ave, Palo Alto, CA 94304

21 Phone: 650-736-8052 Fax 650-725-5671

22 E-mail: nbanaei@stanford.edu

23

24

25 **ABSTRACT**

26 **Background:** Although launched in 2015, little is known about the accuracy of
27 QuantiFERON-TB Gold-Plus (QFT-Plus) for diagnosis of latent *M. tuberculosis* infection
28 (LTBI). Unlike its predecessor, QFT-Plus utilizes two antigen tubes to elicit an immune
29 response from CD4+ and CD8+ T lymphocytes. We conducted a cross-sectional study in
30 low-risk healthcare workers (HCWs) at a single U.S. center to compare QFT-Plus to
31 QuantiFERON-TB Gold in-tube (QFT).

32 **Method:** 989 HCWs were tested with both QFT and QFT-Plus. Risk factors for LTBI were
33 obtained from a questionnaire. QFT-Plus was considered positive if either antigen tube 1
34 (TB1) or TB2 tested positive, per the manufacturer's recommendations, or if both TB1 and
35 TB2 tested positive, using a conservative definition. Results were compared using Cohen's
36 Kappa and linear regression, respectively.

37 **Results:** Agreement of QFT with QFT-Plus was high at 95.6% (95% CI, 94.3-96.9, Kappa,
38 0.57). Majority of discordant results between QFT and QFT-Plus TB1 (84.8%) and QFT and
39 QFT-Plus TB2 (88.6%) fell within the range of 0.2-0.7 IU/mL. Positivity rate in 626 HCWs
40 with no identifiable risk factors and no self-reported history of positive LTBI tests was 2.1%
41 (CI, 1.0-3.2) and 3.0% (CI, 1.7-4.3) with QFT and QFT-Plus, respectively. A conservative
42 definition of QFT-Plus positive yielded a positivity rate of 1.0% (CI, 0.2-1.7, $P = 0.0002$ vs.
43 QFT-Plus and 0.07 vs. QFT). On follow-up testing, of 11 HCWs with discordant QFT-Plus
44 results, 90.9% (10/11) had a negative QFT result.

45 **Conclusions:** The QFT-Plus assay showed high degree of agreement with QFT in U.S.
46 HCWs. A conservative interpretation of QFT-Plus identified nearly all non-reproducible
47 positive results in low-risk HCWs. Larger studies are needed to validate the latter finding
48 and to more clearly define conditions under which a conservative interpretation can be used
49 to minimize non-reproducible positive results in low-risk populations.

50 **Key words:** QFT, QFT-Plus, Healthcare worker, low-incidence, Tuberculosis

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72 **INTRODUCTION**

73 Periodic screening for latent *Mycobacterium tuberculosis* infection (LTBI) is a mandated
74 component of occupation and student health programs in many high-income countries (1). It
75 is intended to identify recently infected individuals and treat them with preventive therapy to
76 avoid development of active disease (2).

77 In the past decade, many health care institutions in the U.S. have switched from tuberculin
78 skin test (TST) to interferon- γ (IFN- γ) release assay (IGRA), in particular the QuantiFERON-
79 TB Gold in-tube assay (QFT, Cellestis/Qiagen, Carnegie, Australia), for annual screening of
80 health care workers (HCWs) (1, 3). Advantages of IGRA over the TST include improved
81 specificity in individuals with bacillus Calmette–Guérin (BCG) vaccination and certain non-
82 tuberculous mycobacterial infections. Moreover, IGRA eliminates the need for a second
83 nurse visit, thus offering operational and economic advantages over TST (2). However,
84 studies conducted in HCWs and students in low-incidence settings have shown high
85 conversion rates with IGRA which exceed the historical or contemporary TST rates (4-
86 6). Also, high rates of reversions and issues with poor reproducibility have also been
87 documented (4, 7). Since positive results can precipitate unnecessary follow up and

88 preventive treatment in low-risk HCWs, the accuracy of IGRA has important implications for
89 patient safety and overutilization of resources (8).

90 In 2015, the next generation of QFT, QuantiFERON-TB Gold-Plus (QFT-Plus) (Qiagen),
91 was launched in Europe and is undergoing clinical trials in the U.S. QFT-Plus employs two
92 TB antigen tubes (TB1 and TB2) for diagnosis of *M. tuberculosis* (MTB) infection. Per the
93 manufacturer's recommendations, QFT-Plus is interpreted positive when either antigen tube
94 results positive. Both antigen tubes include peptides from MTB complex-specific antigens
95 ESAT-6 and CFP-10. While peptides in TB1 and QFT antigen tube are designed to elicit an
96 IFN- γ response from CD4⁺ helper T lymphocytes, TB2 contains an additional set of peptides
97 to also elicit a response from CD8⁺ cytotoxic T lymphocytes. CD8⁺ T lymphocytes are an
98 important component of host immunity to MTB and produce IFN- γ in vitro after stimulation
99 with MTB antigens (9-11). Moreover, ESAT-6- and CFP-10-responsive CD8⁺ T
100 lymphocytes are more frequently detected in subjects with active TB than during latent
101 infection (12-14). They are also detected at a higher frequency after recent infection
102 compared with remote infection (14, 15). Therefore, detection of antigen-responsive CD4⁺
103 and CD8⁺ lymphocytes in QFT-Plus is designed for higher sensitivity in active TB cases and
104 after recent exposure. This was recently suggested in a cohort of 119 patients with active
105 TB (84.9% sensitivity with QFT-Plus TB2 vs. 80.7% with TB1) (16). However, follow-up
106 studies directly comparing QFT to QFT-Plus in active TB patients did not show a difference
107 in sensitivity (15, 17, 18). Furthermore, the sensitivity and specificity of QFT-Plus for
108 LTBI in low-risk individuals such as low-risk North American HCWs remains to be

109 determined. Given the simultaneous availability of two antigen tube results, discordant TB1
110 and TB2 results might indicate false positive results.

111 In a cross-sectional study, we prospectively compared the performance of QFT-Plus to QFT
112 in low-risk HCWs undergoing TB screening at an academic institution in the U.S. We also
113 tested the hypothesis that a more conservative definition of QFT-Plus positivity based on
114 double-positive antigen tube results would reduce positivity rate in low-risk HCWs.

115

116 **METHODS**

117 **Ethics**

118] Per Stanford University Institutional Review Board (IRB), this study was exempt from
119 written informed consent because it constituted a quality improvement project for pre-market
120 validation of QFT-Plus.

121

122 **Study Design**

123 A cross-sectional study was conducted in HCWs at Stanford Health Care to compare the
124 performance of QFT-Plus to QFT and to test the hypothesis that a more conservative
125 interpretation of QFT-Plus results would reduce positivity rate in low-risk HCWs. HCWs
126 with no risk factors and discordant QFT-Plus results were evaluated for LTBI and active
127 tuberculosis (TB) on their follow-up visits.

128

129 **Subjects**

130 Between August 7th and November 19th, 2015, HCWs presenting to the Stanford Health
131 Care Occupational Health Clinic for annual and new employee LTBI screening were
132 randomly enrolled in this study. The occupational health program performs QFT on all
133 HCWs with a negative or undocumented history of LTBI. Risk factors for TB exposure are
134 routinely collected using a questionnaire. HCWs are also evaluated for active TB. LTBI
135 risk factors assessed in a questionnaire include history of close contact with a TB patient,
136 country of birth outside U.S., long-term stay outside the U.S., travel to endemic countries,
137 and employment or volunteer work at high-risk facilities (correctional facility or homeless
138 shelter). Previous history of TST and IGRA positivity and BCG vaccination were also
139 assessed. Positivity rates of QFT and QFT-Plus was assessed in HCWs with no identifiable
140 risk factors and no self-reported history of positive TST or IGRA and also in HCWs with a
141 documented history of negative QFT result in previous year. No-risk HCWs with discordant
142 QFT-Plus results (n = 13) were assessed for active TB and retested with QFT and QFT-Plus
143 on their follow-up visits.

144

145 **QFT and QFT-Plus testing**

146 Blood was drawn, in a single venipuncture, for QFT and QFT-Plus in the following tube
147 order: purge, Nil, QFT-Plus TB1, QFT-Plus TB2, QFT TB Antigen, and Mitogen. Both
148 assays were performed according to the manufacturer's instructions outlined in the package
149 insert. Briefly, blood was drawn into vacutainer tubes up to 1 mL mark and mixed gently.
150 The samples were incubated immediately at 37°C for 16–24 hours and then transported to the
151 clinical microbiology laboratory for ELISA. The plasma was separated by centrifugation and

152 stored at ambient temperature for same-day ELISA or stored at 4°C for ELISA within 72
153 hours. ELISA was performed within 24 hours on an automated robotic ELISA system
154 (DSX; Dynex Technologies, Chantilly, VA). Plasma samples derived from each subject were
155 tested on the same ELISA plate. For QFT, the results are considered positive when TB
156 Antigen minus Nil IFN- γ concentration was ≥ 0.35 IU/mL and $\geq 25\%$ of Nil value. For QFT-
157 Plus, two different interpretative criteria were applied. First, per the manufacturer's
158 instructions, the QFT-Plus assay was interpreted as positive when either TB antigen tube
159 (TB1 or TB2) minus Nil IFN- γ concentration was ≥ 0.35 IU/mL and $\geq 25\%$ of Nil value.
160 Second, using a conservative interpretative criteria (QFT-Plus-C), QFT-Plus was interpreted
161 as positive when both TB antigen tubes (TB1 and TB2) minus Nil IFN- γ concentration were
162 ≥ 0.35 IU/mL and $\geq 25\%$ of Nil value. TB1 and TB2 results were also analyzed separately
163 using the QFT interpretive criteria.

164

165 **Statistical Analysis**

166 Concordance between binary results was measured using Cohen's Kappa (19, 20). Linear
167 regression was used to evaluate quantitative relations between continuous variables. The
168 confidence intervals (CIs) for proportions were calculated from the binomial distribution. The
169 McNemar's test was used to compare proportions. Sample size was calculated as previously
170 described (21). All reported *P* values were two-tailed and calculated with statistical
171 significance set at *P* less than 0.05. Statistical analysis was performed using MedCalc
172 Statistical Software (version 12.3.0, MedCalc Software, Mariakerke, Belgium) and IBM
173 SPSS Statistics 22.0 (IBM Corporation, Armonk, NY, USA).

174

175

176 **RESULTS**177 **Results with QFT and QFT-Plus**

178 In total, 989 HCWs were tested with QFT and QFT-Plus. Demographic data and LTBI risk
179 factors for all HCWs are summarized in Table 1. Two (0.2%) HCWs had indeterminate
180 results with both QFT and QFT-Plus and were therefore excluded from further analysis.
181 Figure 1 summarizes the QFT and QFT-Plus results for the remaining 987 HCWs.
182 Positivity rate with QFT, QFT-Plus, QFT-Plus TB1, and QFT-Plus TB2 was 4.3 % (95% CI,
183 3.0-5.6), 6.4% (CI, 4.9-7.9), 4.2% (CI, 3.0-5.5), and 5.2% (CI, 3.8-6.6), respectively (Table
184 2). Among 31 subjects that tested positive by both QFT and QFT-Plus, 61.3% had one or
185 more risk factors for LTBI. Among 913 subjects that tested negative by both assays, 20.9%
186 had one or more risk factors for LTBI (Supplementary table 1). Among 82 subjects with past
187 history of positive QFT or TST, positivity rate was 20.7% (CI, 11.9-29.5) with QFT and 26.8%
188 (CI, 17.2-36.4) with QFT-Plus. Among 68 subjects with history of BCG vaccination,
189 positivity rate was 17.6% (CI, 8.6-26.7) with QFT and 22.1% (CI, 12.2-32.0) with QFT-Plus.

190

191 **Qualitative and quantitative comparisons between QFT and QFT-Plus**

192 Binary agreement between QFT and QFT-Plus, QFT-Plus TB1, and QFT-Plus TB2 was >95%
193 overall (Table 2 and 3). Agreement in HCWs with one or more risk factors was 90.1% (CI,
194 86.3-94.0), 93.1% (CI, 89.8-96.4), and 93.5% (CI, 90.3-96.7), respectively (Kappa, 0.60, 0.61,
195 and 0.69, respectively). Among 42 (4.3%) HCWs with a positive QFT result, 11 (26.2%)

196 were negative with QFT-Plus (Table 2 and Figure 1). Among 945 (95.7%) HCWs with a
197 negative QFT result, 32 (3.4%) were positive with QFT-Plus (Table 2 and Figure 1). As
198 shown in Figure 2, 84.8% (28/33) and 88.6% (31/35) of HCWs with discordant results
199 between QFT and QFT-Plus TB1 and QFT-Plus TB2, respectively, had a response within the
200 range of 0.2-0.7 IU/mL for one or both assays. Similarly, among 34 HCWs with discrepant
201 results between QFT-Plus TB1 and QFT-Plus TB2, 30 (88.2%) were within the same range.
202 In 626 HCWs with no risk factors for LTBI, 76.9% (10/13) and 81.3% (13/16) of
203 discrepancies between QFT and QFT-Plus TB1 and QFT-Plus TB2, respectively, were within
204 the range of 0.2-0.7 IU/mL (Supplementary figure 1).

205 Quantitative IFN- γ results obtained with QFT showed high degree of correlation with QFT-
206 Plus TB1 and QFT-Plus TB2 (Pearson's correlation coefficient (R) = 0.74 and 0.75,
207 respectively) (Supplementary figure 2). QFT-Plus TB1 and QFT-Plus TB2 also showed very
208 high correlation with each other (R = 0.90). The median TB response in HCWs with positive
209 results was not significantly different between QFT and QFT-Plus TB1 (2.29 vs. 1.77, n=25,
210 $P = 0.21$), QFT and QFT-Plus TB2 (1.58 vs. 1.40, n=29, $P = 1.0$), and QFT-Plus TB1 and
211 TB2 (1.77 vs. 1.89, n=29, $P = 0.29$).

212

213 **Positivity rates with QFT and QFT-Plus in no-risk HCWs**

214 Among 626 HCWs with no identifiable risk factors and no self-reported history of positive
215 TST or IGRA, the positivity rate with QFT and QFT-Plus was 2.1% (CI, 1.0-3.2) and 3.0%
216 (CI, 1.7-4.3), respectively, (Table 4). A more conservative interpretation of QFT-Plus
217 positivity (QFT-Plus-C), based on a double-positive TB1 and TB2 result, yielded a positivity

218 rate of 1.0% (CI, 0.2-1.7) which is significantly lower compared with QFT-Plus ($P < 0.001$)
219 and showed a reduced trend compared with QFT ($P = 0.07$) (Table 4). Among 310 HCWs
220 with a documented history of negative QFT result and no risk factors for LTBI, the positivity
221 rate was 2.6% (CI, 0.8-4.4), 2.6% (CI, 0.8-4.4), and 0.6% (CI, 0-1.5), with QFT, QFT-Plus,
222 QFT-Plus-C, respectively. In this group, the positivity rate with QFT-Plus-C was
223 significantly lower compared with QFT-Plus ($P = 0.03$) and QFT ($P = 0.03$).

224 Follow-up data was available for 11 of the 13 HCWs with discordant QFT-Plus results
225 (Table 5). Zero HCWs developed active tuberculosis during the follow-up period. Ten
226 HCWs had a negative QFT and 6 of 7 HCWs had a negative QFT-Plus (TB1 or TB2),
227 respectively, 9 to 13 months from enrollment. One HCW (study ID No. 6937) who was
228 positive with QFT and QFT-Plus TB2 on enrollment was subsequently positive with QFT
229 after short-term retesting and with QFT and QFT-Plus (TB1 and TB2) 13 months later.
230 Although this HCW did not have any known risk factors for LTBI, he was diagnosed with
231 LTBI and was treated accordingly.

232

233

234 **DISCUSSION**

235 The objective of this study was to compare the performance of QFT-Plus to QFT in low-risk
236 HCWs at a U.S. institution and to test the hypothesis that a more conservative interpretation
237 of QFT-Plus results will reduce positivity rate in HCWs with no known risk factors for LTBI.
238 Overall we found a high degree of agreement (>95%) between QFT-Plus and QFT. We also
239 observed high degree of correlation between quantitative QFT-Plus (TB1 and TB2) and QFT

240 results. For the small number of patients with discordant results, the discordance was mostly
241 due to quantitative results bordering the assay cutoff (0.2-0.7 IU/mL) (22-25). The latter
242 finding is consistent with patterns of discordant results reported in prior IGRA reproducibility
243 studies and suggests that sources of variability previously described for QFT may also be
244 acting on QFT-Plus (25).

245

246 Importantly, in this study we found that 2.1% and 3.0% of 626 HCWs with no identifiable
247 risk factors for LTBI had positive QFT and QFT-Plus results, respectively. The higher
248 positivity rate with QFT-Plus was more frequently due to positive results with TB2 than TB1
249 (2.4% vs. 1.6%). Although IGRAs are more specific than TST in BCG-vaccinated
250 populations (2), IGRAs have proven less specific in low-risk North American HCWs and
251 college students (2). False positive results are likely attributed to one of many sources of
252 variability that cause IGRA results to cross the assay cutoff (25). Because of increasing
253 awareness and recommendations (4, 26), practitioners commonly confirm positive IGRA
254 results in low-risk patients with a short-term follow-up test, which results in added health
255 care costs and overutilization of resources. The QFT-Plus assay, which employs two antigen
256 tubes, was developed for increasing assay sensitivity for active TB by eliciting an IFN- γ
257 response from both CD4+ and CD8+ T lymphocytes. The manufacturer interprets QFT-Plus
258 results as positive when either TB1 or TB2 response reaches the assay cutoff. We showed
259 that a more conservative definition of QFT-Plus positivity, based on double-positive antigen
260 tube results (TB1 and TB2), significantly reduces the positivity rate to 0.6% in risk-free
261 HCWs with a prior negative QFT, which is closer to historical TST conversion rates and

262 somewhat better aligned with the TB epidemiology in the US (4, 7). If we apply the
263 conservative definition to a cohort of 106 low-risk controls from a recent study (16), the
264 positivity rate would drop from 2.8% to 0%. In our study, follow-up investigation of 11 no-
265 risk HCWs with discordant QFT-Plus TB1 and TB2 results showed that in all but one,
266 follow-up testing with QFT and QFT-Plus (TB1 or TB2) remained negative. This finding
267 suggests the conservative interpretation may be a useful strategy for increasing QFT-Plus
268 specificity in low-risk settings. However, larger studies are needed to validate the
269 conservative definition and to better define conditions (i.e., quantitative cutoffs) under which
270 a conservative interpretation can be used to accurately identify non-reproducible positive
271 results in low-risk populations.

272

273 Reproducibility studies have identified different causes of IGRA variability (25). The
274 sources of variability can be broadly classified as pre-analytical, analytical, post-analytical,
275 manufacturing and immunological (25). The use of standardized IGRA testing protocols may
276 minimize variable results after serial testing (25). In this study, QFT and the QFT-Plus assays
277 were simultaneously performed using the same pre-analytical test processes. In this setting,
278 apart from differences in antigen makeup in TB2, analytical variability would have had to
279 arise from ELISA. Metcalfe and colleagues estimated that variability of QFT derived from
280 ELISA is ± 0.6 IU/mL for all individuals and ± 0.24 IU/mL for subjects with initial response
281 in the borderline range of 0.25 to 0.8 IU/mL (23). This is consistent with our study in which
282 most discordant results (84.8% for QFT vs. QFT-Plus TB1 and 88.6% for QFT vs. QFT-Plus
283 TB2) lie in a range of 0.2 to 0.7 IU/mL. The fact that we saw a similar finding in HCWs

284 with no risk factors for LTBI argues for a borderline zone which accounts for variability due
285 to random sources.

286

287 This study had several strengths and weaknesses. The strengths of our study include
288 availability of LTBI risk factors for participants. This allowed us to identify low-risk HCW
289 and calculate positivity rate in this population. In addition, we had prior QFT results for a
290 subset of patients with no risk factors which allowed us to also assess positivity rate in this
291 group. Further, the number of study participants was large and sufficient for comparison of
292 two assays in low-risk HCWs. Lastly, with standardization of pre-analytical processes (25)
293 and simultaneous testing design, we minimized the pre-analytical sources of variability. The
294 fact that all testing was performed under routine clinical practice should render our findings
295 applicable to other health care institutions in non-endemic settings. The limitation of this
296 study includes low number of latently infected HCWs which limited our ability to assess
297 agreement between assays in HCWs with LTBI. It also limited our ability to assess the
298 performance of QFT-Plus in recently exposed versus remotely infected HCWs (14). However,
299 this distribution reflects the low incidence setting we are operating in and therefore our
300 interest to improve assay specificity. Future observational studies in high incidence settings
301 with long term follow ups are needed to assess the sensitivity of QFT-Plus for LTBI after
302 recent exposure.

303

304 In conclusion, the QFT-Plus assay showed high agreement with the QFT assay in low-risk
305 HCW. A conservative interpretation of QFT-Plus identified nearly all positive results in

306 HCW with no known risk factors for LTBI. Larger studies are needed to validate our
307 findings and to better characterize the conservative interpretation in low-risk populations.

308

309 **ACKNOWLEDGMENTS**

310 We thank Qiagen for providing QFT-Plus TB1 and TB2 tubes. Qiagen did not provide any
311 financial support or have any influence on the design and analysis of results in this study.

312

313

314 **REFERENCES**

- 315 1. **Zwerling A, van den Hof S, Scholten J, Cobelens F, Menzies D, Pai M.** 2012.
316 Interferon-gamma release assays for tuberculosis screening of healthcare workers: a
317 systematic review. *Thorax* **67**:62-70.
- 318 2. **Pai M, Denkinger CM, Kik SV, Rangaka MX, Zwerling A, Oxlade O, Metcalfe**
319 **JZ, Cattamanchi A, Dowdy DW, Dheda K, Banaei N.** 2014. Gamma interferon release
320 assays for detection of *Mycobacterium tuberculosis* infection. *Clin Microbiol Rev* **27**:3-20.
- 321 3. **Ringshausen FC, Schablon A, Nienhaus A.** 2012. Interferon-gamma release assays
322 for the tuberculosis serial testing of health care workers: a systematic review. *J Occup Med*
323 *Toxicol* **7**:6.
- 324 4. **Slater ML, Welland G, Pai M, Parsonnet J, Banaei N.** 2013. Challenges with
325 QuantiFERON-TB Gold assay for large-scale, routine screening of U.S. healthcare workers.
326 *Am J Respir Crit Care Med* **188**:1005-1010.

- 327 5. McMullen SE, Pegues DA, Shofer FS, Sheller AC, Wiener EB. 2014.
328 Performance of QuantiFERON-TB Gold and tuberculin skin test relative to subjects' risk of
329 exposure to tuberculosis. *Clin Infect Dis* **58**:1260-1266.
- 330 6. Dorman SE, Belknap R, Graviss EA, Reves R, Schluger N, Weinfurter P, Wang
331 Y, Cronin W, Hirsch-Moverman Y, Teeter LD, Parker M, Garrett DO, Daley CL. 2014.
332 Interferon-gamma release assays and tuberculin skin testing for diagnosis of latent
333 tuberculosis infection in healthcare workers in the United States. *Am J Respir Crit Care Med*
334 **189**:77-87.
- 335 7. Zwerling A, Benedetti A, Cojocariu M, McIntosh F, Pietrangelo F, Behr MA,
336 Schwartzman K, Menzies D, Pai M. 2013. Repeat IGRA testing in Canadian health workers:
337 conversions or unexplained variability? *PLoS One* **8**:e54748.
- 338 8. Menzies D, Joshi R, Pai M. 2007. Risk of tuberculosis infection and disease
339 associated with work in health care settings. *Int J Tuberc Lung Dis* **11**:593-605.
- 340 9. Turner J, Dockrell HM. 1996. Stimulation of human peripheral blood mononuclear
341 cells with live *Mycobacterium bovis* BCG activates cytolytic CD8+ T cells in vitro.
342 *Immunology* **87**:339-342.
- 343 10. Busch M, Herzmann C, Kallert S, Zimmermann A, Hofer C, Mayer D, Zenk SF,
344 Muche R, Lange C, Bloom BR, Modlin RL, Stenger S. 2016. Lipoarabinomannan-
345 responsive Polycytotoxic T Cells are Associated With Protection in Human Tuberculosis. *Am*
346 *J Respir Crit Care Med* **194**:345-355.

- 347 11. **Brookes RH, Pathan AA, McShane H, Hensmann M, Price DA, Hill AV.** 2003.
348 CD8⁺ T cell-mediated suppression of intracellular *Mycobacterium tuberculosis* growth in
349 activated human macrophages. *Eur J Immunol* **33**:3293-3302.
- 350 12. **Rozot V, Vigano S, Mazza-Stalder J, Idrizi E, Day CL, Perreau M, Lazor-**
351 **Blanchet C, Petruccioli E, Hanekom W, Goletti D, Bart PA, Nicod L, Pantaleo G,**
352 **Harari A.** 2013. *Mycobacterium tuberculosis*-specific CD8⁺ T cells are functionally and
353 phenotypically different between latent infection and active disease. *Eur J Immunol* **43**:1568-
354 1577.
- 355 13. **Rozot V, Patrizia A, Vigano S, Mazza-Stalder J, Idrizi E, Day CL, Perreau M,**
356 **Lazor-Blanchet C, Ohmiti K, Goletti D, Bart PA, Hanekom W, Scriba TJ, Nicod L,**
357 **Pantaleo G, Harari A.** 2015. Combined use of *Mycobacterium tuberculosis*-specific CD4
358 and CD8 T-cell responses is a powerful diagnostic tool of active tuberculosis. *Clin Infect Dis*
359 **60**:432-437.
- 360 14. **Nikolova M, Markova R, Drenska R, Muhtarova M, Todorova Y, Dimitrov V,**
361 **Taskov H, Saltini C, Amicosante M.** 2013. Antigen-specific CD4⁻ and CD8⁻ positive
362 signatures in different phases of *Mycobacterium tuberculosis* infection. *Diagn Microbiol*
363 *Infect Dis* **75**:277-281.
- 364 15. **Petruccioli E, Chiacchio T, Pepponi I, Vanini V, Urso R, Cuzzi G, Barcellini L,**
365 **Cirillo DM, Palmieri F, Ippolito G, Goletti D.** 2016. First characterization of the CD4 and
366 CD8 T-cell responses to QuantiFERON-TB Plus. *The Journal of infection* **73**:588-597.
- 367 16. **Barcellini L, Borroni E, Brown J, Brunetti E, Codecasa L, Cugnata F, Dal**
368 **Monte P, Di Serio C, Goletti D, Lombardi G, Lipman M, Rancoita PM, Tadolini M,**

- 369 **Cirillo DM.** 2016. First independent evaluation of QuantiFERON-TB Plus performance. Eur
370 Respir J **47**:1587-1590.
- 371 17. **Hoffmann H, Avsar K, Gores R, Mavi SC, Hofmann-Thiel S.** 2016. Equal
372 sensitivity of the new generation QuantiFERON-TB Gold plus in direct comparison with the
373 previous test version QuantiFERON-TB Gold IT. Clinical microbiology and infection : the
374 official publication of the European Society of Clinical Microbiology and Infectious Diseases
375 **22**:701-703.
- 376 18. **Yi L, Sasaki Y, Nagai H, Ishikawa S, Takamori M, Sakashita K, Saito T,**
377 **Fukushima K, Igarashi Y, Aono A, Chikamatsu K, Yamada H, Takaki A, Mori T,**
378 **Mitarai S.** 2016. Evaluation of QuantiFERON-TB Gold Plus for Detection of
379 *Mycobacterium tuberculosis* infection in Japan. Scientific reports **6**:30617.
- 380 19. **McHugh ML.** 2012. Interrater reliability: the kappa statistic. Biochimica medica
381 **22**:276-282.
- 382 20. **Fleiss JI.** Statistical Methods for Rates and Proportions, 2nd edn. New York: John
383 Wiley & Sons Inc; 1981.
- 384 21. **Banoo S, Bell D, Bossuyt P, Herring A, Mabey D, Poole F, Smith PG, Sriram N,**
385 **Wongsrichanalai C, Linke R, O'Brien R, Perkins M, Cunningham J, Matsoso P,**
386 **Nathanson CM, Olliaro P, Peeling RW, Ramsay A.** 2010. Evaluation of diagnostic tests
387 for infectious diseases: general principles. Nat Rev Microbiol **8**:S17-S29.
- 388 22. **Whitworth WC, Hamilton LR, Goodwin DJ, Barrera C, West KB, Racster L,**
389 **Daniels LJ, Chuke SO, Campbell BH, Bohanon J, Jaffar AT, Drane W, Maserang D,**

- 390 **Mazurek GH.** 2012. Within-subject interlaboratory variability of QuantiFERON-TB gold in-
391 tube tests. *PLoS One* **7**:e43790.
- 392 23. **Metcalfe JZ, Cattamanchi A, McCulloch CE, Lew JD, Ha NP, Graviss EA.** 2013.
393 Test variability of the QuantiFERON-TB gold in-tube assay in clinical practice. *Am J Respir*
394 *Crit Care Med* **187**:206-211.
- 395 24. **Detjen AK, Loebenberg L, Grewal HM, Stanley K, Gutschmidt A, Kruger C,**
396 **Du Plessis N, Kidd M, Beyers N, Walzl G, Hesselning AC.** 2009. Short-term reproducibility
397 of a commercial interferon gamma release assay. *Clin Vaccine Immunol* **16**:1170-1175.
- 398 25. **Banaei N, Gaur RL, Pai M.** 2016. Interferon-gamma release assays for latent
399 tuberculosis: what are the sources of variability? *J Clin Microbiol* **54**:845-850.
- 400 26. **Herrera V, Perry S, Parsonnet J, Banaei N.** 2011. Clinical application and
401 limitations of interferon-gamma release assays for the diagnosis of latent tuberculosis
402 infection. *Clin Infect Dis* **52**:1031-1037.
- 403

404 **FIGURE LEGENDS**

405

406 **Figure 1. Schematic overview of QFT and QFT-Plus results.**

407

408 **Figure 2. Quantitative results in healthcare workers with discordant QFT and QFT-**

409 **Plus results.** Plots show quantitative results for QFT versus QFT-Plus TB1 (A) and QFT

410 versus QFT-Plus TB2 (B) in healthcare workers with discordant results. The dashed reference

411 lines at 0.35 IU/mL are the assay cutoffs and the shaded areas mark the borderline range of

412 0.2–0.7 IU/mL.

413

414

Table 1. Demographic data and LTBI risk factors

Category	n=989
Age in yrs, mean \pm SD	38.0 \pm 11.5
Male gender (%)	301 (30.4%)
Prior positive QFT or TST, No. (%)	82 (8.3%)
BCG vaccination, No. (%)	68 (6.9%)
LTBI risk factor	
0 risk factors, No. (%)	653 (66.0%)
1 risk factor, No. (%)	107 (10.8%)
≥ 2 risk factors, No. (%)	124 (12.6%)
Unknown, No. (%)	105 (10.6%)

QFT, QuantiFERON-TB Gold; TST, tuberculin skin test; BCG, bacillus Calmette–Guérin; LTBI, latent tuberculosis infection

LRBI risk factors per the questionnaire

Table 2. Qualitative comparison between QFT and QFT-Plus

QFT	QFT-Plus		QFT-Plus TB1		QFT-Plus TB2		QFT-Plus-C	
	Positive	Negative	Positive	Negative	Positive	Negative	Positive	Negative
Positive	31 (3.1%)	11 (1.1%)	25 (2.5%)	17 (1.7%)	29 (2.9%)	13 (1.3%)	23 (2.3%)	19 (1.9%)
Negative	32 (3.2%)	913 (92.5%)	16 (1.6%)	929 (94.1%)	22 (2.2%)	923 (93.5%)	6 (0.6%)	939 (95.1%)
Total	63 (6.4%)	924 (93.6%)	41 (4.2%)	946 (95.8%)	51 (5.2%)	936 (94.8%)	29 (2.9%)	958 (97.1%)

QFT, QuantiFERON-TB Gold; QFT-Plus, QuantiFERON-TB Gold-plus; QFT-Plus-C, QuantiFERON-TB Gold-Plus using a conservative interpretation (TB1 and TB2).

Table 3. Qualitative agreement between QFT and QFT-Plus

Comparison	Agreement (%; 95% CI)	Kappa (95% CI)
QFT vs QFT-Plus	944/987 (95.6, 94.3-96.9)	0.57 (0.44-0.70)
QFT vs QFT-Plus TB1	954/987 (96.7, 95.6-97.8)	0.59 (0.45-0.72)
QFT vs QFT-Plus TB2	952/987 (96.5, 95.4-97.7)	0.61 (0.48-0.73)
QFT vs QFT-Plus-C	962/987 (97.4, 96.4-98.4)	0.64 (0.50-0.78)
QFT-Plus TB1 vs QFT-Plus TB2	953/987 (96.6, 95.5-97.7)	0.61 (0.49-0.74)

QFT, QuantiFERON-TB Gold; QFT-Plus, QuantiFERON-TB Gold-Plus; QFT-Plus-C, QuantiFERON-TB Gold-Plus using a conservative interpretation (TB1 and TB2); CI, confidence interval.

Table 4. Positivity rates with QFT and QFT-Plus in 626 healthcare workers with no LTBI risk factors and no prior positive tests

Assay	No. of positives	Positivity rate (95% CI)	<i>P</i> [*]
QFT	13	2.1% (1.0-3.2)	-
QFT-Plus	19	3.0% (1.7-4.3)	0.24
QFT-Plus TB1	10	1.6% (0.6-2.6)	0.58
QFT-Plus TB2	15	2.4% (1.2-3.6)	0.80
QFT-Plus-C [†]	6	1.0% (0.2-1.7)	0.07

QFT, QuantiFERON-TB Gold; QFT-Plus, QuantiFERON-TB Gold-Plus; QFT-Plus-C, QuantiFERON-TB Gold-Plus using a conservative interpretation (TB1 and TB2), ^{*}Compared with QFT using McNemar's test, [†]*P*= 0.0002 compared with that of QFT-Plus

Table 5. Follow-up results for 13 no-risk healthcare workers with discordant QFT-Plus results.

Study No.	Age (yr)	Sex (M/F)	Enrollment Result				Follow-up Result						
			QFT		QFT-Plus		QFT		QFT-Plus		Since last screen		
			Initial screen	Short-term retest	TB1	TB2	Annual screen	Short-term retest	TB1	TB2	Interval (mo)	TB exposure	Active TB
6937	53	M	0.4	0.44	0.27	0.77	1.01	ND	0.91	1.12	13	No	No
823	30	M	0.47	0.16	0.34	0.36	0.16	ND	ND	ND	12	No	No
907	28	F	1.47	0.02	0.5	0.05	0.03	ND	ND	ND	13	No	No
1716	38	F	0.06	ND	0.45	0.25	0.16	ND	0.21	0.25	12	No	No
3958	28	F	0.07	ND	1.85	0.14	0	ND	0.03	0.01	13	No	No
6258	28	F	0.02	ND	5.11	0.02	0	ND	ND	ND	10	No	No
3720	26	F	0	ND	0	1.26	0	ND	0.13	0.15	13	No	No
4749	58	F	0	ND	0	0.67	0	ND	0.00	0.34	12	No	No
885	34	F	0.06	ND	0.23	0.62	0.03	ND	0.01	0.17	9	No	No
6156	23	F	0	ND	0.04	0.60	ND	ND	ND	ND	NA	NA	NA
2262	51	M	0.01	ND	0.06	0.48	0.01	ND	0.01	0.03	11	No	No
1588	55	M	0.28	ND	0.23	0.36	0.6	0.15	ND	ND	12	No	No
4698	43	F	0	ND	0.01	0.35	ND	ND	ND	ND	NA	NA	NA

TB Ag minus Nil (IU/mL) are shown and positive results are shaded.

ID no., identification number; yr, year; M, male; F, female, mo, month; TB, tuberculosis; ND, not done; NA, not available

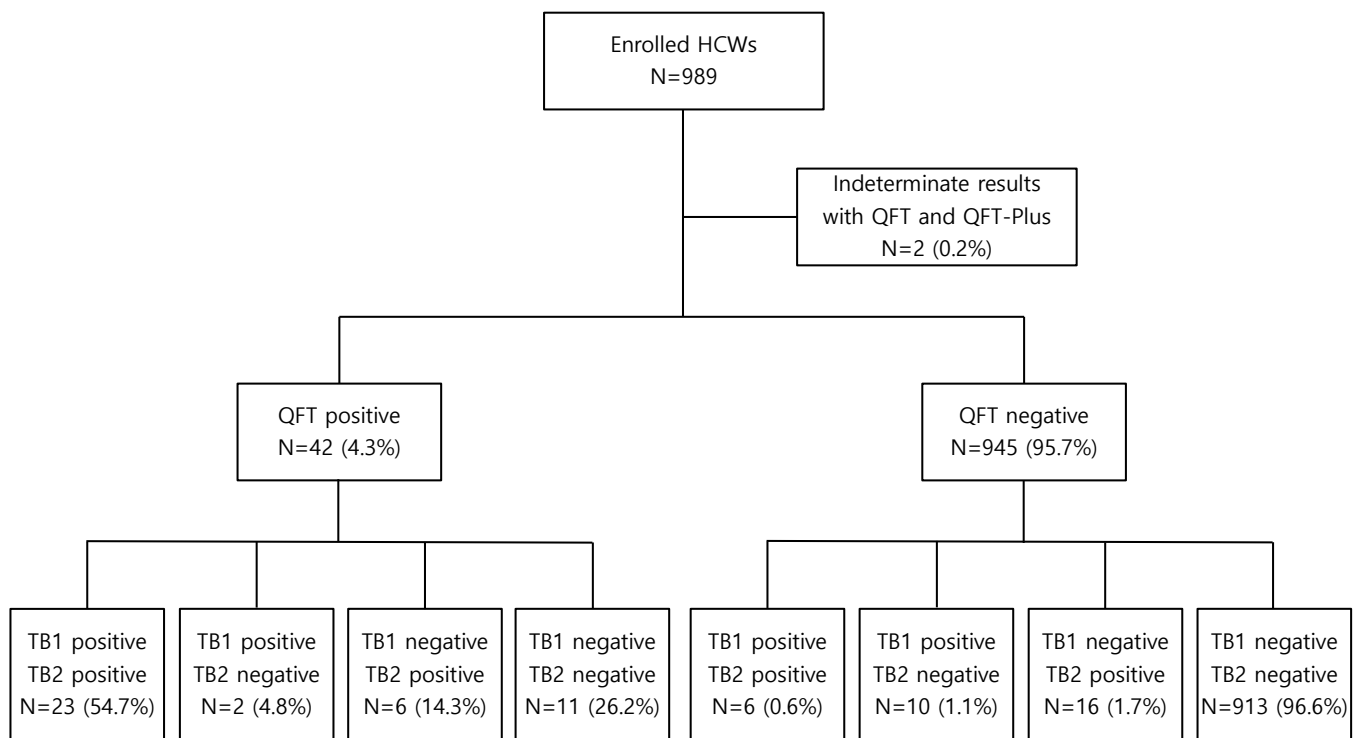


Figure 1. Schematic overview of QFT and QFT-Plus results.

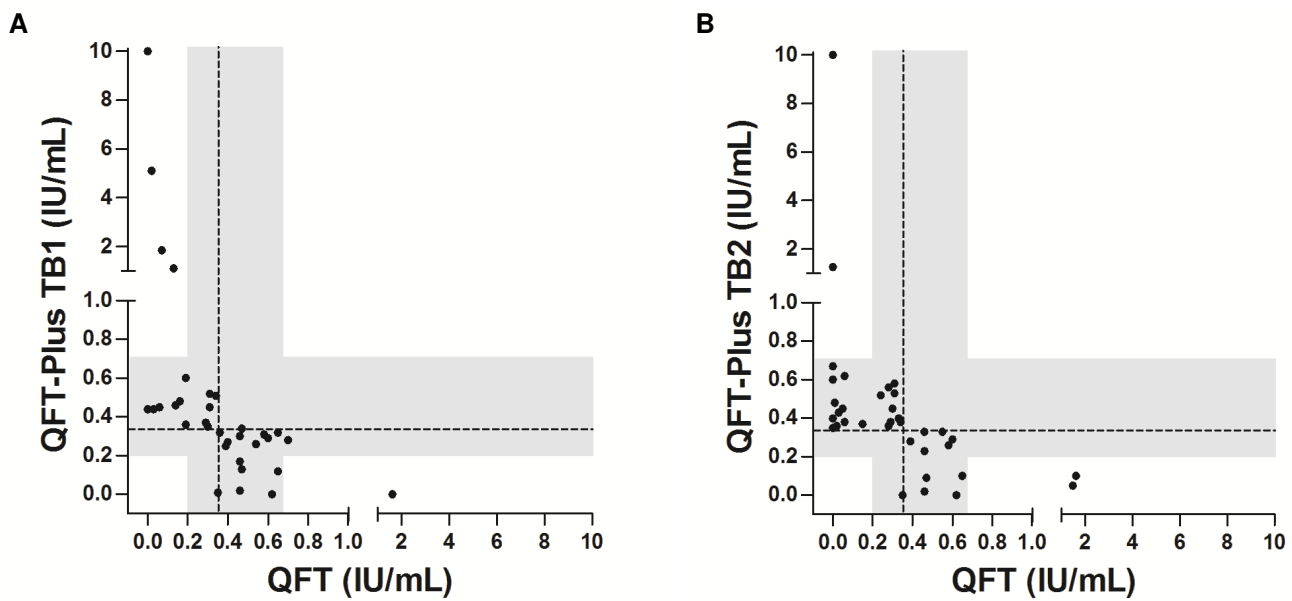


Figure 2. Quantitative results in healthcare workers with discordant QFT and QFT-Plus results. Plots show quantitative results for QFT versus QFT-Plus TB1 (A) and QFT versus QFT-Plus TB2 (B) in healthcare workers with discordant results. The dashed reference lines at 0.35 IU/mL are the assay cutoffs and the shaded areas mark the borderline range of 0.2–0.7 IU/mL.